# Somatic evolutionary genomics: Mutations during development cause highly variable genetic mosaicism with risk of cancer and neurodegeneration

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Somatic mutations must happen often during development because of the large number of cell divisions to expand from a single-cell zygote to a full organism. A mutation in development carries forward to all descendant cells, causing genetic mosaicism. Widespread genetic mosaicism may influence diseases that derive from a few genetically altered cells, such as cancer. I show how to predict the expected amount of mosaicism and the variation in mosaicism between individuals. I then calculate the predicted risk of cancer derived from developmental mutations. The calculations show that a significant fraction of cancer in later life likely arises from developmental mutations in early life. In addition, much of the variation in the risk of cancer between individuals may arise from variation in the degree of genetic mosaicism set in early life. I also suggest that certain types of neurodegeneration, such as amyotrophic lateral sclerosis (ALS), may derive from a small focus of genetically altered cells. If so, then the risk of ALS would be influenced by developmental mutations and the consequent variation in genetic mosaicism. New technologies promise the ability to measure genetic mosaicism by sampling a large number of cellular genomes within an individual. The sampling of many genomes within an individual will eventually allow one to reconstruct the cell lineage history of genetic change in a single body. Somatic evolutionary genomics will follow from this technology, providing new insight into the origin and progression of disease with increasing age.

amyotrophic lateral sclerosis | Luria–Delbruck fluctuation analysis | somatic phylogenetics

A human develops from a single cell. From that single cell, an individual grows to  $10^{13}$  to  $10^{14}$  cells. That growth requires many cell divisions and, consequently, much somatic mutation must occur during development. Those developmental mutations likely have significant consequences for genetic mosaicism in the body and for the risk of cancer that arises from those mutations (1, 2).

In this article, I will show how to predict the amount of somatic mutation and the amount of genetic variability in the body. I will connect those calculations to the risk of cancer. I also propose that certain neurodegenerative diseases that occur later in life may often derive from early-life somatic mutations that occur during development.

Before turning to the details, let us consider in a general way the magnitude of somatic mutation during development in relation to the number of cells in the body. During development the single-cell zygote expands to  $N = 10^{13}$  to  $10^{14}$  cells. How many cell divisions occur during that expansion? Each time a cell divides, the number of cells in the body increases by one, assuming no cell death. So, to start with one cell and expand to N cells requires at least N - 1 cell divisions.

How much somatic mutation occurs during development? We do not have good measurements, but we can make some rough calculations. The minimum number of cells divisions is  $N - 1 \approx N = 10^{13}$  to  $10^{14}$ . Define the mutation rate per gene per cell division as u. No truly reliable estimates of somatic mutation

rates exist, but typically assumed values are of the order  $u = 10^{-7}$  to  $10^{-6}$  (3). The total number of mutational events per gene during development is the mutation rate per cell division multiplied by the number of cell divisions,  $uN = 10^{6}$  to  $10^{8}$ . Thus, every gene in the genome mutates many times.

The value of uN measures the number of mutational events that occur in each gene. But most often, we will be interested in the number of cells that carry a mutation in a particular gene. For example, if a mutation occurs early in development, then that single mutation will carry forward to many descendant cells. By contrast, relatively few cells will carry a mutation that happens late in development. To understand the relation between the number of mutations that occur and the number of cells that carry a mutation, we must place somatic mutations in the context of cell lineage history. In other words, we must think of the body in relation to the lineage history descending from the single ancestral zygote and how mutations accumulate in that lineage history.

The accumulation of change within the lineage history of the body is somatic evolutionary genomics. With  $\approx 10^{13}$  to  $10^{14}$  cells in a body, and probably  $> 10^{16}$  cells produced over a lifetime, the lineage history within a single individual is much greater than for all of the hominids that have ever lived, perhaps as great as for all of the primates that have ever lived.

The tremendous evolutionary history within each human body has, until recently, been hidden by the difficulty of measuring genetic changes in cells. New high-throughput genomic technologies are just opening up the possibility of directly measuring somatic variability and evolution (4). To understand the evolutionary history of the individual and the consequences for disease, we must place somatic genomics within the context of the rate and pattern of evolutionary change in cellular lineages.

#### **Genetic Mosaicism**

In this section, I explain in more detail how mutations accumulate in cell lineages. I emphasize that the shape of lineage history differs at different times of life and in different tissues, affecting the patterns of somatic evolution. I also show how to predict the amount of genetic mosaicism in an individual and the variation in mosaicism between individuals. The following sections connect the amount and variation in genetic mosaicism to the risk of diseases such as cancer and neurodegeneration.

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**Fig. 1.** Lineage history of cells in renewing tissues. All cells trace their ancestry back to the zygote. Each tissue, or subset of tissue, derives from a precursor cell;  $n_p$  rounds of cell division separate the precursor cell from the zygote. From a precursor cell,  $n_e$  rounds of cell division lead to exponential clonal expansion until the descendants differentiate into the tissue-specific stem cells that seed the developing tissue. In a compartmental tissue, such as the intestine, lineage history of the renewing tissue follows an essentially linear path, in which each cellular history traces back through the same sequence of stem cell divisions (2, 21). At any point in time, a cell traces its history back through  $n_s$  stem cell divisions to the ancestral stem cell in the tissue, and  $n = n_p + n_e + n_s$  divisions back to the zygote. (Modified from figure 13.1 in ref. 2, based on the original in ref. 1.)

**Shape of Cell Lineages.** Fig. 1 shows that renewing tissues typically have two distinct phases in the history of their cellular lineages. Early in life, cellular lineages expand exponentially to form the tissue. For the remainder of life, stem cells renew the tissue by dividing to form a nearly linear cellular history

Mutations accumulate differently in the exponential and linear phases of cellular division (1). During the exponential phase of development, a mutation carries forward to many descendant cells. During the renewal phase, a mutation transmits only to the localized line of descent in that tissue compartment: one mutational event has limited consequences (Fig. 2).

Amount and Variation of Genetic Mosaicism: Theory. Consider a renewable tissue, such as the colon epithelium or the hematopoietic system. Those tissues renew throughout life from a set of stem cells. A human colon has approximately  $N = 10^8$  stem cells, with probably at least that many stem cells in the hematopoietic system. At the end of development, what fraction of those stem cells carries a somatically derived mutation? To answer this question, we must analyze how mutations during development translate into the number of initially mutated stem cells at the end of development.

Mutations occur stochastically in the small number of cells present early in development. The number of mutant stem cells and the degree of genetic mosaicism will therefore vary greatly between individuals according to a probability distribution called the Luria–Delbruck distribution (5). That distribution describes the number of mutant cells, M, in a population that grows exponentially from one cell to N cells (6, 7).

Suppose, for example, that  $N = 10^8$  stem cells must be produced during development to seed a tissue. Exponential growth of one cell into N cells requires, in the absence of cell death, a total of N - 1 cellular divisions arranged into approximately  $\ln(N)$  cellular generations. In this case,  $\ln(10^8) \approx 18$ . If the mutation rate per locus per cell division during exponential growth is  $u_e$ , then the probability, x, that any final stem cell carries a mutation at a particular locus is approximately the mutation rate per cell division multiplied by the number of cell divisions from that particular cell back to the ancestral progenitor cell. In this case,  $x = u_e \ln(N)$ . This probability is usually small: for example, if  $u_e = 10^{-6}$ , then x is of the order of  $10^{-5}$ .

The frequency of initially mutated stem cells may be small, but the number may be significant. The average number of mutated cells at a particular locus is the number of cells, N, multiplied by the probability of mutation per cell, x. In this example,  $Nx \approx 10^3$ , or  $\approx 1,000$ .



**Fig. 2.** Mutational events in development occur at different stages in the exponential, branching phase of cellular expansion. In this example, the red mutation happens early, causing a significant fraction of somatic cells to carry the same mutation by descent. By contrast, the green mutation happens late in development, causing only a small fraction of somatic cells to carry the same mutation by descent. Many mutations will arise within the stem cells, each stem cell renewing only a very small fraction of all somatic cells. For example, the blue mutation is private to a single stem cell and will be confined to the small subset of somatic cells derived from that stem cell. Branching lines represent the developmental phase of cellular expansion, the large cells are stem cells, and the small cells in each line form a clone derived from their stem cell ancestor.



**Fig. 3.** Number of initially mutated stem cells at the end of development. The *N* initial stem cells derive by exponential growth from a single precursor cell. Each plot shows the cumulative probability, *p*, for the number of mutated initial stem cells. By plotting  $\log_{10}(p/(1-p))$ , the zero line gives the median of the distribution. The number above each line is  $u_{e_r}$  the mutation probability per cell added to the population during exponential growth. (I used an actual value of  $10^{-5.2}$  rather than  $10^{-5}$  because of computational limitations.) For a single gene, the mutation probability per gene per cell division,  $u_g$ , is probably  $>10^{-7}$ . If there are at least L = 100 genes for which initial mutations can influence the progression to cancer, then  $u_e = Lu_g \ge 10^{-5}$ . Initial mutations may, for example, occur in DNA repair genes, causing an elevated rate of mutation at other loci. Calculations were made with algorithms in Zheng (38). (Modified from figure 13.3 in ref. 2, based on the original in ref. 1.)

I have focused on mutations at a single locus. Mutations at many different loci may predispose to genetically influenced diseases such as cancer. Suppose mutations at L different loci can contribute to predisposition. We can get a rough idea of how multiple loci affect the process by simply adjusting the mutation rate per cell division to be a genome-wide rate of predisposing mutations, equal to  $u_e L$ . The number of loci that may affect predisposition may reasonably be around  $L \approx 10^2$  and perhaps higher. Following the calculation in the previous paragraph, with  $L \geq 10^2$ , the number of initial stem cells carrying a predisposing mutation would on average be at least  $10^5$ . Some individuals might have two predisposing mutations in a single initial stem cell.

The average number of initially mutated cells tells only part of the story, because the distribution for the number of mutants is highly skewed. A few rare individuals have a great excess; in those individuals, a mutation arises early in development, and most of the stem cells would carry the mutation. Those individuals would have nearly the same risk as one who inherited the mutation.

Fig. 3 shows the distribution for the number of initially mutated stem cells at the end of development. For example, in Fig. 3 *Right*, with a mutation probability per cell division of  $10^{-6}$ , a *y*-axis value of 2 means that  $\approx 10^{-2}$ , or 1%, of the population has  $>10^4$  initially mutated stem cells at a particular locus (L = 1). Similarly, with a mutation probability per cell division of  $10^{-7}$ , a *y*-axis value of 3 means that  $\approx 10^{-3}$ , or 0.1%, of the population has  $>10^4$  initially mutated stem cells at a particular locus.

Amount and Variation of Genetic Mosaicism: Observations. My simple calculations show that, in a typical individual, every gene mutates somatically many times. Similarly, most cells in the body probably carry at least one somatic mutation. When we place this widespread somatic mosaicism into the context of cell lineage history, as in Fig. 2, we see that the body likely comprises variable-sized patches of somatic mosaicism throughout the genome.

Simple theory tells us that this widespread mosaicism must be present. But few measurements of mosaicism have been accomplished. The lack of measurement occurs because it is not easy to analyze genetic variation between individual cells in the huge population of cells that comprise a single body. I mentioned earlier that the population of cells in a single body greatly exceeds that of all hominid individuals that have ever existed. A comprehensive study of the somatic evolutionary genomics of a single body would be as challenging as an evolutionary analysis of genetic variability for all humans and their hominid ancestors throughout hominid history.

A few analyses of mosaicism do exist (8). Mutation of a skin pigmentation gene in development causes skin cells to be marked by the mutation. The marked cells trace the tips of the somatic evolutionary lineage tree on the body surface. Interestingly, the patterned skin markings vary considerably. For example, several visible skin diseases follow the lines of Blaschko, which trace out what seem like contour lines or whorls over the skin surface (9–12). Other distinct patterns also occur in skin diseases (12). Speckled lentiginous naevus and Becker's naevus follow a mosaic checkerboard pattern; mosaic trisomy of chromosome 13 causes scattered leaf-like shapes of hypopigmentation.

Although somatic evolutionary genomics is difficult at present, genomic technology is advancing rapidly. The recent cancer genome project shows the potential for screening genetic changes in the somatic cells of an individual (4). Wallace's (13) work and future vision for somatic mitochondrial genetics emphasize the potential for analyzing the diseases of increasing age in the context of accumulating somatic mutations in cellular lineages.

The hematopoietic system provides a particularly promising somatic component for future study. Blood cells derived from diverse stem cell populations can easily be sampled and followed over time. Greaves (14) reviewed several lines of evidence demonstrating that developmental mutations in the hematopoietic system cause widespread somatic mosaicism. In many cases, those developmental mutations appear to be the primary cause of childhood leukemia (15–17).

Are the rare childhood forms of leukemia isolated examples, or is somatic mosaicism derived from developmental mutation a hidden risk factor in many cancers? At present, no direct evidence answers this question. The following section considers some theory by which we can predict the risk from developmental mutation. The theory also provides a framework for analyzing the data on somatic genomics that will become available in the future.

### **Risk of Cancer**

Developmental mutations inevitably cause genetic mosaicism. Those cells carrying somatic mutations from development may be predisposed to cancer. In this section, I analyze the increased risk of cancer attributable to somatic mutations in early development.

**Frank and Nowak's Model.** Fig. 3 shows the probability distribution for the number of stem cells that have mutations. The number of mutated stem cells is M = Nz, where N is the total number of stem cells, and z is the frequency of those stem cells that carry a mutation that predisposes to disease. In this section, I focus on the average frequency of mutated stem cells, where x is the average of z, which leads to the average number of mutated stem cells, Nx. In the following section, I discuss the wide variation between individuals in the frequency of mutated stem cells. That variation in mutated stem cells may explain a significant fraction of the variation in cancer risk between individuals.

Mutations during the exponential phase of cellular growth in development cause the average frequency of stem cells with mutations to be  $x \approx u_e \ln(N)$  (6), where  $u_e$  is the mutation rate during exponential cellular growth in development. Although the frequency of stem cells that start with a mutation may be small, those mutations can contribute substantially to the total risk of cancer. Suppose, by the multistage model of cancer progression, that k rate-limiting mutations are needed to cause

cancer (18–20). To obtain a rough estimate of the total risk, let  $R_T = N(1 - x)R_k + NxR_{k-1}$ , where x is the frequency of stem cells that start with one mutation, and  $R_k$  is the risk that a particular stem cell lineage acquires k mutations during the phase of linear division and tissue renewal. The risk,  $R_k$ , can be approximated by the  $\gamma$  distribution, which gives the probability for the occurrence of the kth event over a particular time interval. From the  $\gamma$  distribution,  $R_k \approx (u_s \tau)^k / k!$ , where  $u_s$  is the mutation rate per stem cell division, and  $\tau$  is the total number of stem cell divisions.

The relative odds that cancer derives from stem cells that begin with one mutation acquired in development compared with those stems cells that lack a developmental mutation can be calculated as  $F = xR_{k-1}/(1 - x)R_k \approx u_e \ln(N)k/u_s\tau$ . The next step is to fill in approximate magnitudes for these quantities. We can take  $k \approx 5$  for the number of rate-limiting mutations to cause epithelial cancer in humans (18–20),  $\ln(N) \approx 20$ , and  $\tau$  from the range 100 to 1,000. This gives the relative odds of cancer deriving from a developmental mutation, *F*, ranging from  $\approx u_e/10 u_s$  to  $u_e/u_s$ .

If mutations accumulate with the same probability per cell division during exponential growth and linear stem cell division,  $u_e = u_s$ , then mutations arising in development increase risk by 10–100%. If, as Cairns (21) has argued, stem cell mutation rates are much less than mutation rates during exponential growth,  $u_s \ll u_e$ , then almost all cancer arises from predisposed stem cell lineages that were mutated during development.

**Meza et al.'s Model.** Meza et al. (22) extended Frank and Nowak's (1) model in three ways. First, they used an explicit quantitative model of colorectal cancer progression to apply direct parameter estimates from data. Second, they analyzed the relative proportion of cancer derived from developmental mutations at each age; they showed that earlier onset of cancer more often derives from developmental mutations than later onset of cancer. Third, they calculated the amount of variation in cancer risk between individuals caused by the stochastic nature of somatic mutations in early development.

Meza et al. (22) began with a model of colorectal cancer progression and incidence that they had previously studied (23). In that model, carcinogenesis progresses through four stages: two initial transitions, followed by a third transition that triggers clonal expansion, and then a final transition to the malignant stage.

In their study, Meza et al. (22) began with the same four-stage model. They then added a Luria–Delbruck process to obtain the probability distribution for the number of stem cells mutated at the end of development. The stochasticity in the Luria–Delbruck process causes wide variation between individuals in the number of mutated stem cells. Meza et al. first calculated the probability that an individual carries Nz initially mutated stem cells at the end of development. To obtain overall population incidence, they summed the probability for each Nz multiplied by the incidence for individuals with Nz mutations.

Meza et al. (22) summed incidence in their four-stage model over the number of initially mutated stem cells to fit the model's predicted incidence curve to the observed incidence of colorectal cancer in the United States. From their fitted model, they then estimated the proportion of cancers attributable to mutations that arise during development. Fig. 4 shows that a high proportion of cancers may arise from mutations during the earliest stage of life.

Cancers at unusually young ages are often attributed to inheritance. However, Fig. 4 suggests that early-onset cancers may often arise from developmental mutations. Developmental mutations act similarly to inherited mutations: if the developmental mutation happens in one of the first rounds of postzygotic cell division, then many stem cells start life with the mutation.



**Fig. 4.** The proportion of cancers that arise from cells mutated during development. These plots show calculations based on a specific four-stage model of colorectal cancer progression (22). The parameters of the progression model were estimated from incidence data. The values of *u* above each plot show the mutation rate per year in stem cells. Itsely divide between 10 and 100 times per year, thus a mutation rate per year of at least  $10^{-5}$  per locus seems reasonable. In each plot, the three curves sketch the heterogeneity between individuals in risk attributable to developmental mutations. The first quartile shows the proportion of cancers at each age for those individuals whose risk is in the lowest 25% of the population, in particular, those individuals who by chance have the fewest stem cells mutated during development. Similarly, the fourth quartile shows the risk for the highest 25% of the population with regard to developmental mutations. [Reproduced with permission from ref. 22 (Copyright 2005, Elsevier).]

Inheritance is, in effect, a mutation that happened before the first zygotic division.

#### **Risk of Neurodegenerative Disease**

Developmental mutations alter a fraction of cells, leading to genetic mosaicism. Any disease that derives from a small number of mutated cells may be influenced by developmental mutations. For example, cancer progression ultimately encompasses many cells of a tissue, but the initial change in one or a few cells starts the process. Thus, developmental mutations, by seeding the process in many stem cells, enhance the risk that one of those predisposed cells will lead to cancer.

Neurodegenerative diseases such as amyotrophic lateral sclerosis (ALS) appear, at first glance, very different from cancer. Individuals typically seem to be without any symptoms through middle or late life. Then, within a few years, they progress from initial motor problems to widespread motor neuron degeneration and death (24). The widely distributed motor neuron degeneration seems to argue against the disease initiating in a one or a few cells. However, Armon (25) suggested that the disease does start in a focal area and then spreads to neighboring motor neurons.

In ALS, a particular limb may show the first signs of motor neuron deterioration. The next symptoms frequently occur in the contralateral limb at the same spinal level and at contiguous spinal levels (26). Alternatively, spread may occur up and down the motor neuron system through the primary motor neurons directly connected to muscles and those neurons that feed into the primary motor system (27). Focal initiation followed by spread means that the disease may be initiated in one or a few cells, in the same way as cancer (25).

Developmental mutations and somatic mosaicism would play a role in ALS risk only if the initiating events in the focal cells derived from genetic (or epigenetic) changes in those cells. No direct evidence exists for a role of somatic mutation in initiating ALS. However, inherited germ-line mutations do strongly predispose to ALS, so it is possible that genetics plays a key role in disease (27–32). But the direct role of somatic mutation, and of what follows, remains a speculative hypothesis at present.

The most common genetic mutations associated with ALS lead to misfolded proteins (33–35). Individuals who inherit a predisposing mutation typically appear normal until disease onset usually in the age range of 25–65 years (30). As in inherited predisposition to cancer, other factors must be involved in transforming predisposed cells into the initiating focus of a disease that subsequently spreads.

Most cases seem to arise sporadically, without any evidence of inherited predisposition. Sporadic cases typically have a later age of onset in the range 40–80 years (27). The sporadic disease is mostly similar to the familial form, although some variations in the inherited form have been noted (27). This similarity between sporadic and inherited cases suggests that the sporadic cases may possibly begin from an initiating focus of cells with genetic mutations, but, again, there is no direct evidence for this.

If both inherited and sporadic cases do initiate disease from a small focus of cells that carry a genetic mutation, then, as in cancer, we can predict a continuum of risk. Those who inherit a predisposing mutation have the highest risk and earliest age of onset, with all cells carrying the mutation. Those who suffer a somatic mutation early in development have only slightly lower risk and later age of onset relative to inherited cases, with many cells carrying the mutation. Those who suffer a developmental mutation at some intermediate time in neuronal development have less risk and higher age of onset, with fewer cells, but still a significant number, carrying the predisposing mutation. Those who suffer a late developmental mutation have relatively low risk and late age of onset, with relatively few cells carrying the mutation. Finally, those with very few developmentally mutated cells may form the largest class with lowest risk and latest onset; those with very few developmental mutations may in fact have such low risk that nearly all cases can be ascribed to those who carry a significant number of developmental mutations.

If most cases derive from either inherited or developmental mutations, then the risk of ALS and perhaps other neurodegenerative diseases may be set very early in life. If sporadic cases often derive from developmental mutations, then somatic genomics will reveal an association between age of onset and the fraction of cells that carry a predisposing somatic mutation. The inherited cases would simply be the extreme of the risk continuum, in which all cells carry the predisposing mutation.

What sort of evidence would weigh in favor or against this speculative hypothesis? On the positive side: new somatic genomic measures that show a correlation between early-onset ALS and a somatic mutation widely distributed in neural tissue by descent from a single somatic mutation in a common ancestral cell; a mechanistic pathway that links the somatic mutation to the onset and progression of ALS; and further evidence that disease spreads from a small focus of cells that have been transformed. On the negative side: no correlation between early-onset ALS and widely distributed somatic mutations known to predispose to ALS when those mutations arise in the germ line; and evidence that disease arises over a short period in many widely distributed locations rather than spreading from a small number of foci.

#### **Potential Difficulties**

The risks conferred by abundant low-penetrance genes affecting carcinogenesis may be of the same order as the risk from developmental mutations. How can we pick out the proportion of cancers that arise from developmental mutations if they are "drowned out" by population heterogeneity because of a number of weakly penetrant germ-line susceptibility genes?

We can distinguish between the risk caused by developmental mutations and weakly penetrant germ-line mutations by direct evidence from somatic genomics. Evidence favors developmental mutations when: one directly observes that somatic mutations dispersed widely in a tissue arose by normal cell lineage expansion in the development of that tissue; the dispersed somatic mutation derived from a single mutational event in an ancestral cell; and there is an association between those somatic mutations from normal developmental processes and the probability and age of cancer onset. These sorts of measurements are not easy at present, but the technology is developing very rapidly.

Another potential difficulty concerns the simple multistage model of carcinogenesis that I used in making calculations about the consequences of developmental mutations. I used the classical and most commonly cited form of the multistage model presented by Armitage and Doll (18) in 1954. As early as 1957, Armitage and Doll (36) pointed out that the age-specific incidence of certain cancers did not fit the simplest assumptions of their original multistage theory. There has followed a long history of model variants that fit particular assumptions to the age-specific incidence of particular cancers (2).

Recently, Meza et al. (37) made an important contribution to this long history by fitting explicit models of carcinogenesis to the age-specific incidence of colorectal and pancreatic cancers. They show that the classical model of Armitage and Doll (18) is not sufficient, and they develop a specific alternative that accounts for certain aspects of the biology and provides a much better fit to the data.

Given the limitations of the classical Armitage and Doll model that I used in the early sections of this article, how might my conclusions about developmental mutations be affected by alternative and more realistic models of carcinogenesis? The particular quantitative values derived from the model and shown in the figures would be changed. However, the model and the figures were meant only to illustrate the basic qualitative idea that developmental mutation potentially contributes to the risk of cancer and potentially alters the age of onset in those who suffer most strongly from widely dispersed developmental mutations. Those qualitative conclusions hold under any realistic model of carcinogenesis in which somatic mutations (or any heritable changes to cells) play a role.

My recent book (2) went into great detail about the various quantitative models of carcinogenesis in relation to the available data on age-specific incidence. Much can be learned by close study of those issues, and Meza et al. (37) have made a very significant contribution to that topic. But with regard to developmental mutations and somatic genomics, the real problems turn on the coming technological advances that will allow direct observation of how somatic mutations disperse through tissues in relation to cell lineage history and how those somatic mutations influence progression to cancer. Confidence in particular mathematical models must await those advances.

## Conclusions

Skin diseases and childhood leukemia show that somatic mosaicism does occur. I have argued that those examples are just hints of the hidden and widespread mosaicism that arises from developmental mutations. Commonly accepted assumptions about cell division, cell lineage history, and somatic mutation lead inevitably to the prediction that mosaicism is common in all genes throughout the genome.

Importantly, the stochastic nature of mutation and the small number of cells in early development predict that the degree of mosaicism varies greatly between individuals. For any gene, a small fraction of individuals in the population will carry a somatic mutation in many cells, the mutation having occurred in early development. Those individuals will be at risk for diseases such as cancer and neurodegeneration that, later in life, can spread from a small focus of genetically predisposed cells.

Other individuals will carry relatively few predisposing somatic mutations derived from development. Those individuals have less risk later in life for diseases such as cancer and

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neurodegeneration. If this argument is correct, then a significant fraction of risk for diseases later in life may derive from mutational events before birth.

Until recently, genetic technology has not allowed widespread sampling of somatic genomes within an individual. Thus, there is currently no direct evidence for or against this theory of widespread mosaicism and its association with the risk of disease. New technologies promise the ability to sample large numbers of genomes. The sampling of many genomes within an individual will eventually allow one to reconstruct the cell lineage history of genetic change in a single body. Somatic evolutionary genomics will follow from this technology, providing new insight into the origin and progression of disease with increasing age.

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